

## EXTERNAL REFERENCES

### ID SCREEN® CHLAMYDIA ABORTUS INDIRECT MULTI-SPECIES

Last update: December 2025

#### Publications / References:

#### PERFORMANCE EVALUATION

|   |   |                                    |  |  |  |  |
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| <p>1)Mermoud I. <i>et al.</i> (2017). <b>Comparison of two ELISA tests to study the seroprevalence of <i>CHLAMYDIA abortus</i> in the New Caledonian bovine herd.</b> Poster presented at WALVD, Sorrento (Italy), June 7-10, 2017.</p> | <ul style="list-style-type: none"> <li>prospective study of the seroprevalence of <i>CHLAMYDIA abortus</i> on 790 <b>bovine</b> sera from a <i>Chlamydia abortus</i>-free area using the IDEXX Chlamydiosis Total Ab Test. The ID SCREEN® CHLAMYDIA ABORTUS Indirect Multi-species ELISA was used to confirm positive results obtained with the IDEXX kit. Sera confirmed positive were then tested by CFT.</li> <li><b>Results:</b> -using the Iddex test: 33% of the sera were interpreted positive or doubtful (n=256)<br/>-using the ID SCREEN® CHLAMYDIA ABORTUS Indirect Multi-species: only 11 positive and 4 doubtful sera<br/>-all sera were CFT negative</li> </ul> <p><b>The results obtained with the ID SCREEN® CHLAMYDIA ABORTUS Indirect Multi-species match the epidemiological situation of chlamydial infection in ruminants in the studied area, with a high specificity., unlike the other commercial test.</b></p> | <p>Comparison with competitors</p> | <p>Correlation with other techniques</p> |  |  |  |
| <p>2)Schuber E. <i>et al.</i> (2015). <b>Ring trial of the German Reference Laboratory for chlamydiosis.</b> Poster presented at the AVID Meeting, Kloster Banz (Germany), September 9-11, 2015.</p>                                    | <ul style="list-style-type: none"> <li>ring trial: 20 serum samples, including 11 <b>sheep</b> sera from a herd showing abortions or from a vaccination study, and 9 <b>cattle</b> sera from a herd without clinical signs were tested using the ID SCREEN® CHLAMYDIA ABORTUS Indirect Multi-species, IDEXX Chlamydiosis Total Ab Test and CFT.</li> <li><b>Trial conclusion: -the ID SCREEN® CHLAMYDIA ABORTUS Indirect Multi-species is specific for <i>C. abortus</i> without unspecific cross reactions with other chlamydial species and correctly identified all <i>C. abortus</i>-positive sera.</b></li> </ul> <p>-using the Iddex test, besides <i>C. abortus</i>, antibodies to other <i>chlamydia</i> species were also detected</p>   | <p>Comparison with competitors</p> | <p>Correlation with other techniques</p> |  |  |  |

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|  | <p>-CFT cannot be recommended for diagnosis of chlamydial infections.</p>  |                                    |  |  |  |  |
| <p>3)Horigan M.W <i>et al.</i> (2009). <b>CHLAMYDIA abortus – An Evaluation of Three Commercial ELISAs.</b> Poster presented at WAVLD Madrid (Spain), June 18-20, 2009.</p>  | <ul style="list-style-type: none"> <li>comparative study of 3 commercial ELISAs (including the ID SCREEN® CHLAMYDIA ABORTUS Indirect Multi-species=kit B) and comparison with CFT.</li> <li><b>Results: the study demonstrates the superior specificity and overall high performances of the ID SCREEN® CHLAMYDIA ABORTUS Indirect Multi-species:</b> <ul style="list-style-type: none"> <li>-specificity: 96.2-100%</li> <li>-sensitivity: 70%</li> <li>-high correlation with CFT (97.4% CFT-negative samples were negative with the ID SCREEN® CHLAMYDIA ABORTUS Indirect Multi-species.</li> </ul> </li> </ul> | <p>Comparison with competitors</p> | <p>Correlation with other techniques</p> |  |  |  |
| <p>4)Marques P.X. <i>et al.</i> (2008). <b>A comparison of three serological methods for the identification of pregnant ewes infected with CHLAMYDIA abortus.</b> Poster presented at WALVD, Dublin (Ireland), 2008.</p> | <ul style="list-style-type: none"> <li>comparative study of 2 commercial ELISAs (the ID SCREEN® CHLAMYDIA ABORTUS Indirect Multi-species and another commercial test, no longer available) with immunoblot.</li> <li><b>Results:</b> relative to the immunoblot assay:           <ul style="list-style-type: none"> <li>-using the ID SCREEN® CHLAMYDIA ABORTUS Indirect Multi-species: 95.7% specificity and 54.2% sensitivity.</li> <li>-using the other test: 95.2% specificity and 48.1% sensitivity.</li> </ul> </li> </ul>   | <p>Comparison with competitors</p> | <p>Correlation with other techniques</p> |  |  |  |
| <p>5)Pourquier P. <i>et al.</i> (2007). <b>Preliminary validation of a new commercial ELISA kit for the detection of antibodies directed against C. abortus.</b> Poster presented at the WAVLD Conference, 2007.</p>     | <ul style="list-style-type: none"> <li>-comparative study of the ID SCREEN® CHLAMYDIA ABORTUS Indirect Multi-species, CFT and <i>C. pecorum</i> immunofluorescence test.</li> <li><b>Results:</b> -no or low cross-reactions with <i>C. pecorum</i> <ul style="list-style-type: none"> <li>-high sensitivity: all sera with a CFT titre superior or equal to 1/20 were found positive, with an excellent correlation was obtained between CFT titres and S/P values.</li> </ul> </li> </ul>  |                                    | <p>Correlation with other techniques</p> |  |  |  |

## EPIDEMIOLOGICAL STUDIES

### LARGE RUMINANTS

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|---|---|--|--|--|--|--|--|
| <p>6)Yahiaoui F. <i>et al.</i> (2025). <b>Seroprevalence of Abortion-Related infectious diseases and associated risk factors among Brucellosis-free herds in Northern central Algeria.</b> BMC Veterinary Research, 21(1), 185.</p>   | <ul style="list-style-type: none"> <li>• 132 <b>cow</b> sera were tested using the ID SCREEN® CHLAMYDIA ABORTUS Indirect Multi-species.</li> <li>• <i>Results:</i> seroprevalence was 4.1%.</li> </ul>  |  |  |  |  |  |  |
| <p>7)Liang L. <i>et al.</i> (2021). <b>Seroprevalence of <i>Chlamydia abortus</i> infection in yak (<i>Bos grunniens</i>) in Tibet, China.</b> Irish Veterinary Journal, 74(1), 19.</p>   | <ul style="list-style-type: none"> <li>• 938 <b>yak</b> sera were tested using the ID SCREEN® CHLAMYDIA ABORTUS Indirect Multi-species.</li> <li>• <i>Results:</i> seroprevalence was 11.1%.</li> </ul>   |  |  |  |  |  |  |
| <p>8)Kandel R. <i>et al.</i> (2020). <b>Seroprevalence of <i>Chlamydia abortus</i> Among Cattle of Selected Areas in Terai Belt of Nepal.</b> International Journal of Applied Sciences and Biotechnology, 8(3), 363-367.</p>   | <ul style="list-style-type: none"> <li>• blood samples from 92 <b>cattle</b> displaying signs or history of anestrus were tested using the ID SCREEN® CHLAMYDIA ABORTUS Indirect Multi-species.</li> <li>• <i>Results:</i> seroprevalence was 2.17%.</li> </ul>                                   |  |  |  |  |  |  |
| <p>9)Djellata N. <i>et al.</i> (2020). <b>Prevalence and factors associated with a higher or lower risk of exposure to <i>Coxiella burnetii</i>, <i>Chlamydia abortus</i> and <i>Toxoplasma gondii</i> in dairy cows that have aborted in Algeria.</b> Revue Scientifique et Technique. Office International des Epizooties, 38(3).</p> | <ul style="list-style-type: none"> <li>• 368 sera from <b>cattle</b> that had aborted on 124 farms were tested using the ID SCREEN® CHLAMYDIA ABORTUS Indirect Multi-species.</li> <li>• <i>Results:</i> seroprevalence was<br/>-at animal-level: 12.23%<br/>-at herd-level: 29.84%.</li> </ul>   |  |  |  |  |  |  |
| <p>10)Pokharel S. <i>et al.</i> (2020). <b>Seroprevalence of <i>Chlamydia abortus</i> in Anestrus Cattle of Nawalpur and Chitwan District, Nepal.</b> Nepalese Veterinary Journal, 29-41.</p>   | <ul style="list-style-type: none"> <li>• blood samples from 92 <b>cattle</b> displaying signs or history of anestrus were tested using the ID SCREEN® CHLAMYDIA ABORTUS Indirect Multi-species.</li> <li>• <i>Results:</i> seroprevalence was 2.17%.</li> </ul>                                   |  |  |  |  |  |  |
| <p>11)Majed R. <i>et al.</i> (2018). <b>Preliminary study of seroprevalence of <i>CHLAMYDIA abortus</i> amongst cattle in Ninavah province.</b> Adv. Anim. Vet. Sci. 6(3): 135-138.</p>   | <ul style="list-style-type: none"> <li>• sera from 368 cows (150 <b>aborted cows</b>, 150 <b>pregnant cows</b>, and 68 <b>calves</b> aged 1 month old) were tested using the ID SCREEN® CHLAMYDIA ABORTUS Indirect Multi-species.</li> <li>• <i>Results:</i> seroprevalence was 0.82%.</li> </ul> |  |  |  |  |  |  |

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| <p>12)Shrestha P. (2016). <b>Seroprevalence of CHLAMYDIA abortus in Aborted and Infertile Dairy Cattle in Chitwan District, Nepal.</b> Nepalese Veterinary Journal, 11.</p> | <ul style="list-style-type: none"> <li>• 92 sera from <b>cows</b> with history of reproductive disorders such as abortion, repeat breeding and anestrus were tested using the ID SCREEN® CHLAMYDIA ABORTUS Indirect Multi-species.</li> <li>• <i>Results:</i> seroprevalence was 7.6%.</li> </ul> |  |
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## SMALL RUMINANTS

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| <p>13)Baloğlu H. <i>et al.</i> (2025). <b>Preliminary seroprevalence study of zoonotic abortigenic agents in the abortion inexperienced sheep population in the Northern Cyprus.</b> Veterinary Research Communications, 49(4), 219.</p>                                 | <ul style="list-style-type: none"> <li>• 450 <b>sheep</b> sera collected from 45 farms were tested using the ID SCREEN® CHLAMYDIA ABORTUS Indirect Multi-species.</li> <li>• <i>Results:</i> seroprevalence was<br/>-at animal-level: 3.11%<br/>-at herd-level: 17.77%.</li> </ul> |  |
| <p>14)Kifouli A.H. <i>et al.</i> (2025). <b>Enzootic Ovine Abortion among small ruminants in Southern Benin.</b> Veterinaria Italiana, 61(1).</p>  | <ul style="list-style-type: none"> <li>• sera from 200 <b>sheep</b> and 185 <b>goats</b> were tested using the ID SCREEN® CHLAMYDIA ABORTUS Indirect Multi-species.</li> <li>• <i>Results:</i> seroprevalence was<br/>-in sheep: 7.62%<br/>-in goats: 5.14%.</li> </ul>            |  |
| <p>15)Lei F. <i>et al.</i> (2025). <b>Seroprevalence of Chlamydia abortus infection in Tibetan sheep in Qinghai Province, China.</b> Acta Tropica, 264, 107593.</p>  | <ul style="list-style-type: none"> <li>• 1043 <b>sheep</b> sera were tested using the ID SCREEN® CHLAMYDIA ABORTUS Indirect Multi-species.</li> <li>• <i>Results:</i> seroprevalence was 14.7%.</li> </ul>   |  |
| <p>16)Sağlam A G. <i>et al.</i> (2025). <b>An investigation on the potential role of Q fever and chlamydiosis of ovine abortion.</b> Journal of Advances in VetBio Science and Techniques, 10(1), 35-41.</p>   | <ul style="list-style-type: none"> <li>• blood samples from 100 aborted <b>sheep</b> were tested using the ID SCREEN® CHLAMYDIA ABORTUS Indirect Multi-species.</li> <li>• <i>Results:</i> 5 sera tested positive.</li> </ul>  |  |
| <p>17)Tarusikirwa D.F. <i>et al.</i> (2023). <b>Seroprevalence and assessment of public awareness of Brucella spp., Toxoplasma gondii and Chlamydia abortus in small ruminants from selected smallholder commercial farms of Zimbabwe.</b> PLoS ONE 18(6): e0287902.</p> | <ul style="list-style-type: none"> <li>• sera from 335 <b>goats</b> and 63 <b>sheep</b> were tested using the ID SCREEN® CHLAMYDIA ABORTUS Indirect Multi-species.</li> <li>• <i>Results:</i> seroprevalence was<br/>-in goats: 9.9%<br/>-in sheep: 4.8%.</li> </ul>               |  |

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| <p>18)Zeeshan M.A. <i>et al.</i> (2023). <b>Seroepidemiological study of zoonotic bacterial abortifacient agents in small ruminants.</b> <i>Front. Vet. Sci.</i> 10: 1195274.</p>   | <ul style="list-style-type: none"> <li>• sera from 235 <b>goats</b> and 150 <b>sheep</b> were tested using the ID SCREEN® CHLAMYDIA ABORTUS Indirect Multi-species.</li> <li>• <i>Results:</i> seroprevalence was<br/>-in goats: 4.6%<br/>-in sheep: 3.3%.</li> </ul>   |  |  |  |  |  |
| <p>19)Nankam R.C. <i>et al.</i> (2022). <b>Seroprevalence and Risk Factors Associated with <i>Toxoplasma gondii</i> and <i>CHLAMYDIA abortus</i> Infection in Domestic Small Ruminants in Cameroon.</b> <i>Parasitologia</i>, 2, 198–205.</p> | <ul style="list-style-type: none"> <li>• 1061 <b>sheep</b> sera from 200 flocks were tested using the ID SCREEN® CHLAMYDIA ABORTUS Indirect Multi-species.</li> <li>• <i>Results:</i> seroprevalence was<br/>-at flock-level: 3%<br/>-at individual level: 4.2%.</li> </ul>   |  |  |  |  |  |
| <p>20)Zurovac-Sapundžić Z. <i>et al.</i> (2022). <b>Seroprevalence of <i>CHLAMYDIA abortus</i> in sheep in the Belgrade epizootiological area during 2019-2021.</b> <i>Archives of Veterinary Medicine</i>, 15(1), 85-92.</p>                 | <ul style="list-style-type: none"> <li>• 552 <b>sheep</b> sera were tested using the ID SCREEN® CHLAMYDIA ABORTUS Indirect Multi-species.</li> <li>• <i>Results:</i> seroprevalence was 6%.</li> </ul>  |  |  |  |  |  |
| <p>21)Iraninezhad Z. <i>et al.</i> (2020). <b>Seroepidemiological feature of <i>Chlamydia abortus</i> in sheep and goat population located in northeastern Iran.</b> In <i>Veterinary Research Forum</i> (Vol. 11, No. 4, p. 423).</p>        | <ul style="list-style-type: none"> <li>• 271 <b>sheep</b> and 181 <b>goat</b> sera samples from 40 sheep/goat epidemiologic units were tested using the ID SCREEN® CHLAMYDIA ABORTUS Indirect Multi-species.</li> <li>• <i>Results:</i> seroprevalence was<br/>-at animal-level: 9.7%<br/>-at epidemiologic unit-level: 70%.</li> </ul> |  |  |  |  |  |
| <p>22)Mamlouk A. <i>et al.</i> (2020). <b>Seroprevalence and associated risk factors of <i>Chlamydia abortus</i> infection in ewes in Tunisia.</b> <i>Comparative Immunology, Microbiology and Infectious Diseases</i>, 71, 101500.</p>       | <ul style="list-style-type: none"> <li>• 803 <b>ewe</b> sera from 26 herds were tested using the ID SCREEN® CHLAMYDIA ABORTUS Indirect Multi-species.</li> <li>• <i>Results:</i> seroprevalence was:<br/>-at animal-level: 6.6%<br/>-at herd-level: 58%.</li> </ul>   |  |  |  |  |  |
| <p>23)Salman S.S. <i>et al.</i> (2020). <b>Serological and biochemical study of ovine chlamydiosis in Baghdad city.</b> <i>Plant Archives</i> Vol. 20, No. 1, pp. 1926-1929.</p>  | <ul style="list-style-type: none"> <li>• sera from 14 <b>rams</b> and 146 <b>ewes</b> were tested using the ID SCREEN® CHLAMYDIA ABORTUS Indirect Multi-species.</li> <li>• <i>Results:</i> seroprevalence was<br/>-in ewes: 2.55%<br/>-in rams: 28.57%.</li> </ul>   |  |  |  |  |  |
| <p>24)Borujeni M.P. <i>et al.</i> (2019). <b><i>Chlamydia abortus</i> infection in goats in the southwest of Iran.</b> <i>Revue de medecine veterinaire</i>, 1(170), 9-14.</p>  | <ul style="list-style-type: none"> <li>• 368 goat sera were tested using the ID SCREEN® CHLAMYDIA ABORTUS Indirect Multi-species.</li> <li>• <i>Results:</i> seroprevalence was 5.71%.</li> </ul>   |  |  |  |  |  |

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| <p>25)Fahad O.A. <i>et al.</i> (2017). <b>Survey for ovine and caprine chlamydiosis by ELISA in AL-Fallujah city/Iraq.</b> Journal of Entomology and Zoology Studies, 5(6), 322-326.</p>                       | <ul style="list-style-type: none"> <li>sera from 124 <b>ewes</b>, 30 <b>rams</b>, 21 <b>does</b>, and 9 <b>bucks</b> were tested using the ID SCREEN® CHLAMYDIA ABORTUS Indirect Multi-species.</li> <li><b>Results:</b> seroprevalence was:           <ul style="list-style-type: none"> <li>-in ewes: 9.67%</li> <li>-in rams: 3.33%</li> <li>-in does: 28.57%</li> <li>-in bucks: 22.22%.</li> </ul> </li> </ul> |  |  |  |  |  |
| <p>26)Jalboush N. <i>et al.</i> (2017). <b>Detection of CHLAMYDIA abortus antibody in active reproductive rams in sheep herds in northern Palestine.</b> Revue de Medecine Veterinaire, 168(7-9), 192-196.</p> | <ul style="list-style-type: none"> <li>2806 <b>ram</b> sera from 353 farms were tested using the ID SCREEN® CHLAMYDIA ABORTUS Indirect Multi-species.</li> <li><b>Results:</b> seroprevalence was           <ul style="list-style-type: none"> <li>-at animal-level: 13.7%</li> <li>-at herd-level: 53.3%.</li> </ul> </li> </ul>   |  |  |  |  |  |
| <p>27)Villagra-Blanco R. <i>et al.</i> (2015). <b>Detection of antibodies against CHLAMYDIA abortus in Costa Rican sheep flocks.</b> Open veterinary journal, 5(2), 122-126.</p>                               | <ul style="list-style-type: none"> <li>359 <b>sheep</b> sera from 15 flocks were tested using the ID SCREEN® CHLAMYDIA ABORTUS Indirect Multi-species.</li> <li><b>Results:</b> seroprevalence was           <ul style="list-style-type: none"> <li>-at flock-level: 5.29%</li> <li>-at animal-level: 19%.</li> </ul> </li> </ul>   |  |  |  |  |  |

## CAMELIDS

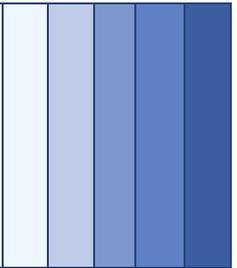
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| <p>28)Selim A. <i>et al.</i> (2024). <b>Serosurvey and associated risk factors for Chlamydia abortus infection in Dromedary camels in Egypt.</b> Tropical Animal Health and Production, 56(5), 188.</p>                                    | <ul style="list-style-type: none"> <li>410 camel sera were tested using the ID SCREEN® CHLAMYDIA ABORTUS Indirect Multi-species.</li> <li><b>Results:</b> seroprevalence was 6.6%.</li> </ul>  |  |  |  |  |  |
| <p>29)Selmi R. <i>et al.</i> (2024). <b>Serological and molecular survey of brucellosis and chlamydiosis in dromedary camels from Tunisia.</b> Comparative Immunology, Microbiology and Infectious Diseases, 104, 102098.</p>              | <ul style="list-style-type: none"> <li>470 camel sera were tested using the ID SCREEN® CHLAMYDIA ABORTUS Indirect Multi-species.</li> <li><b>Results:</b> seroprevalence was 5.75%.</li> </ul> |  |  |  |  |  |
| <p>30)Benaissa M.H. <i>et al.</i> (2020). <b>First report of CHLAMYDIA abortus infection in the dromedary camel (Camelus dromedarius) population in eastern Algeria.</b> Comparative Immunology, Microbiology and Infectious Diseases.</p> | <ul style="list-style-type: none"> <li>865 camel sera were tested using the ID SCREEN® CHLAMYDIA ABORTUS Indirect Multi-species.</li> <li><b>Results:</b> seroprevalence was 2.5%.</li> </ul>  |  |  |  |  |  |

## WILDLIFE

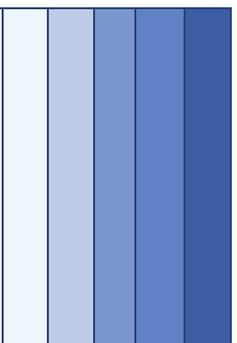
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| <p>31)Jiménez-Martín D. <i>et al.</i> (2025). <b>Seroepidemiological study of <i>Chlamydia abortus</i> in domestic and wild ruminants in Spanish Mediterranean ecosystems.</b> Preventive Veterinary Medicine, 106600.</p> | <ul style="list-style-type: none"> <li>• sera from small ruminants (390 <b>sheep</b> and 390 <b>goats</b>), and free-ranging wild ruminants (390 <b>red deer</b> (<i>Cervus elaphus</i>), 110 <b>mouflon</b> (<i>Ovis aries musimon</i>) and 105 <b>Iberian ibex</b> (<i>Capra pyrenaica</i>)) were tested using the ID SCREEN® CHLAMYDIA ABORTUS Indirect Multi-species.</li> <li>• <b>Results:</b> seroprevalence was           <ul style="list-style-type: none"> <li>-in goats: 34.1 %</li> <li>-in sheep: 22.8 %</li> <li>-in roe deer: 1.8 % of red deer</li> <li>-in Iberian ibex: 3.8 %</li> <li>-in mouflon: 4.6 %.</li> </ul> </li> </ul> |  |
| <p>32)Žele Vengušt D. <i>et al.</i> (2024). <b>Seroprevalence of infectious pathogens of zoonotic and veterinary importance in wild ruminants from Slovenia.</b> Front. Vet. Sci. 11:1415304.</p>                          | <ul style="list-style-type: none"> <li>• 312 sera from wild ruminants (134 <b>roe deer</b>, 113 <b>red deer</b>, 53 <b>Alpine chamois</b>, 10 <b>European mouflon</b>, and 2 <b>Alpine ibexes</b> were tested using the ID SCREEN® CHLAMYDIA ABORTUS Indirect Multi-species.</li> <li>• <b>Results:</b> overall seroprevalence was 0.96%.</li> </ul>  |  |
| <p>33)Tavernier P. <i>et al.</i> (2015). <b>Serologic screening for 13 infectious agents in roe deer (<i>Capreolus capreolus</i>) in Flanders.</b> Infection ecology &amp; epidemiology, 5(1), 29862.</p>                  | <ul style="list-style-type: none"> <li>• 178 sera from <b>roe deer</b> were tested using the ID SCREEN® CHLAMYDIA ABORTUS Indirect Multi-species.</li> <li>• <b>Results:</b> seroprevalence was 6.7%.</li> </ul>  |  |

## HORSES

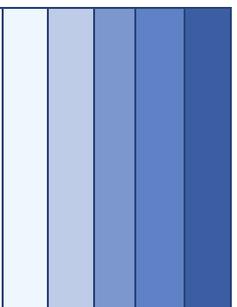
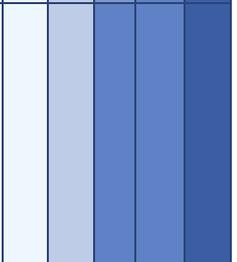
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| <p>34)Rubio-Navarrete I. <i>et al.</i> (2017). <b>Prevalence of <i>Chlamydia abortus</i> antibodies in horses from the northern state of Mexico and its relationship with domestic animals.</b> Journal of equine veterinary science, 56, 110-113.</p> | <ul style="list-style-type: none"> <li>• sera from 301 <b>horses</b>, 25 <b>bovines</b>, 8 <b>goats</b>, and 94 <b>sheep</b> were tested using the ID SCREEN® CHLAMYDIA ABORTUS Indirect Multi-species.</li> <li>• <b>Results:</b> seroprevalence was           <ul style="list-style-type: none"> <li>-in horses: 1.32%</li> <li>-in bovines: 48%</li> <li>-in goats: 12.5%</li> <li>-in sheep: 29.7%.</li> </ul> </li> </ul> |  |
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| <p>35)Jiménez D. <i>et al.</i> (2014). <b>Serosurveillance of infectious agents in equines of the Central Valley of Costa Rica.</b> Open Veterinary Journal, 4(2), 107-112.</p> | <ul style="list-style-type: none"> <li>• 181 <b>equine</b> sera were tested using the ID SCREEN® CHLAMYDIA ABORTUS Indirect Multi-species.</li> <li>• <b>Results:</b> seroprevalence was 4.8%.</li> </ul> |  |
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## HUMANS

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| <p>36)Kifouly A.H. <i>et al.</i> (2025) <b>First assessment of the health status of pregnant women, detection of prevalence and risk factors associated with enzootic ovine abortion disease among pregnant women in southern Benin.</b> Front. Public Health 13:1532390.</p> | <ul style="list-style-type: none"> <li>• sera from 385 <b>pregnant humans</b> were tested using the ID SCREEN® CHLAMYDIA ABORTUS Indirect Multi-species.</li> <li>• <b>Results:</b> seroprevalence was 1.3% (5 positive cases).</li> </ul> |  |
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## VACCINATION/EXPERIMENTAL STUDIES

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| <p>37)Murcia-Belmonte A. <i>et al.</i> (2019). <b>Effect of progesterone on the vaccination and immune response against <i>Chlamydia abortus</i> in sheep.</b> Veterinary immunology and immunopathology, 213, 109887.</p> | <ul style="list-style-type: none"> <li>• experimental study in <b>sheep</b> immunized using an experimental inactivated <i>C. abortus</i> vaccine then challenged with <i>C.abortus</i>; seroconversion was followed using the ID SCREEN® CHLAMYDIA ABORTUS Indirect Multi-species.</li> <li>• <b>Results:</b> sheep of the vaccinated groups manifested seroconversion after the first vaccination at 28 days post-vaccination.</li> </ul> |  |
| <p>38)Destrez A. <i>et al.</i> (2017). <b>Effects of a chronic stress treatment on vaccinal response in lambs.</b> animal, 11(5), 872-880.</p>   | <ul style="list-style-type: none"> <li>• experimental study in <b>lambs</b> exposed to stress and infected using <i>Chlamydia abortus</i> attenuated vaccine strain 1B; seroconversion was followed using the ID SCREEN® CHLAMYDIA ABORTUS Indirect Multi-species.</li> <li>• <b>Results:</b> at the days 3, 7, 14 and 21 post-vaccination, stress did not have an effect on vaccination.</li> </ul>  |  |

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| <p>39)García-Seco T. <i>et al.</i> (2016). <b>Effect of Preventive <i>Chlamydia abortus</i> Vaccination in Offspring Development in Sheep Challenged Experimentally.</b> <i>Front. Vet. Sci.</i> 3:67.</p>      | <ul style="list-style-type: none"> <li>• study of the efficacy in <b>ewes</b> of an inactivated standard commercial vaccine (SV= standard dose) and a 1/2 diluted dose (DV=1/2 dose) in pregnant sheep challenged with <i>C. abortus</i>; seroconversion was followed using the ID SCREEN® CHLAMYDIA ABORTUS Indirect Multi-species.</li> <li>• <b>Results:</b> ewes from the control group (non-vaccinated) remained negative until the challenge, while positive responses were recorded at 29 days post vaccination (first sampling post-vaccination) in both SV and DV groups; after the challenge, the highest proportion of reactors in the 3 groups (SV, DV and Control) was observed at 14 days post-challenge.</li> </ul>   |  |
| <p>40)Álvarez D. <i>et al.</i> (2015). <b>Intratracheal infection as an efficient route for testing vaccines against <i>Chlamydia abortus</i> in sheep.</b> <i>The Veterinary Journal</i>, 205(3), 393-398.</p> | <ul style="list-style-type: none"> <li>• experimental vaccination on <b>sheep</b> using an inactivated <i>C. abortus</i> vaccine, a challenge experimentation with 2 different infection pathways (intranasal IN and intratracheal IT) was then applied using <i>C. abortus</i> strain AB7; seroconversion was followed using the ID SCREEN® CHLAMYDIA ABORTUS Indirect Multi-species.</li> <li>• <b>Results:</b> the experimental infection elicited a marked specific humoral response in all cases, although the response was much more rapid and higher in the vaccinated groups; on the day of inoculation (day 0), all vaccinated animals presented high mean seropositivity. This mean level had increased by 8 days post-infection (dpi) and remained high until 22 dpi; non-vaccinated animals did not seroconvert until 8 dpi in the case of IT-animals and until 22 dpi in the case of IN animals.</li> </ul> |  |

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