

## EXTERNAL REFERENCES

# ID SCREEN® RIFT VALLEY FEVER COMPETITION MULTI-SPECIES

Last update: November 2025

### Publications / References:

#### PERFORMANCE STUDIES

1)Dafalla O.M. <i>et al.</i> (2025). <b>Integrated Assessment of Antibody Responses to RVFV Using Competitive ELISA and VNT in Vaccinated Animal Samples from Southwest Saudi Arabia.</b> Researchsquare.	<ul style="list-style-type: none"> <li>20 <b>sheep</b> serum samples were collected one-month post-vaccination (with the RVF Smithburn live attenuated vaccine); the antibody response to Rift Valley fever virus was evaluated via viral neutralization tests (<b>VNT</b>), virus titration and the ID Screen RIFT VALLEY FEVER Competition Multi-species.</li> <li><b>Results:</b> -ID Screen: antibody detection was heavily identified at serum dilutions of 1:2 – 1:32 with S/N 3%-23%.</li> <li><b>-strong correlation between VNT neutralization at low dilutions and ELISA positivity.</b></li> </ul>	Correlation with other techniques				
2)Hungerbuhler V. <i>et al.</i> (2024). <b>Diagnostic serology test comparison for Q fever and Rift Valley fever in humans and livestock from pastoral communities.</b> PLoS Negl Trop Dis 18(10): e0012300.	<ul style="list-style-type: none"> <li>this study aimed to assess the agreement of test results for RVF in <b>humans</b> (91 sera) and <b>livestock</b> (102 sera) across different laboratory conditions and, for humans, different types of diagnostic tests (ID Screen RIFT VALLEY FEVER Competition Multi-species and <b>IFA</b>).</li> <li><b>Results:</b> variability in inter-laboratory diagnostic test agreement for RVF serology in humans and livestock; test agreements ranged from fair to moderate (Cohen's kappa) or almost perfect considering PABAK.</li> </ul>	Correlation with other techniques				Interlaboratory assay
3)Milićević V. <i>et al.</i> (2024). <b>Evaluation of commercial ELISA kits' diagnostic specificity for FAST diseases in wild animals.</b> Onderstepoort Journal of Veterinary Research 91(1), a2164.	<ul style="list-style-type: none"> <li>evaluation of the specificity of the ID Screen RIFT VALLEY FEVER Competition Multi-species and another commercial ELISA kit (RVF Ingezim, Ingenasa) using 342 serum samples from <b>wild ruminants</b> and 100 serum samples from <b>wild boar</b>; sera were tested twice, then a third test was performed after serum inactivation.</li> <li><b>Results:</b> -specificity of the ID Screen was: 98.83% (first test) and 98.83% (second test) before serum inactivation 99.71% after serum inactivation</li> </ul>	Comparison with competitors				Specificity data

	- agreement with the Ingenasa kit was 98.8%.					
4)Pedarrieu A. <i>et al.</i> (2021). <b>External quality assessment of Rift Valley fever diagnosis in countries at risk of the disease: African, Indian Ocean and Middle-East regions.</b> Plos one, 16(5), e0251263.	<ul style="list-style-type: none"> <li>serological inter-laboratory proficiency test: the ID Screen RIFT VALLEY FEVER Competition Multi-species, another commercial kit, and two in-house serological assays for the detection of RVFV-specific IgG antibodies were tested. Out of the 18 laboratories that participated in the PT, 14 used the ID Screen RIFT VALLEY FEVER Competition Multi-species. The analytical performance of test sensitivity and specificity based on the <b>seroneutralization test</b> considered as the reference was 100%. The panel consisted of 20 samples of sera including negative (n = 7) and positive sera (n = 11), as well as sera at the limit of detection (n = 2).</li> <li><b>Results:</b> -13 out of 14 laboratories that used the ID Screen RIFT VALLEY FEVER Competition Multi-species reported correct results for all the samples regarding the criteria of sensitivity, repeatability, specificity, and dose-response relationship compared to SNT -the 13 laboratories that used the ID Screen RIFT VALLEY FEVER Competition Multi-species reported 100% correct results (Cohens 'Kappa value = 1) whereas a kappa value of 0.88 was reported for the other commercial kit used by three laboratories. -the results obtained by the three labs that use the other commercial kit showed a lower sensitivity than the ID Screen RIFT VALLEY FEVER Competition Multi-species. <b>The ID Screen RIFT VALLEY FEVER COMPETITION MULTI-SPECIES showed excellent performance, better than the other commercial kit.</b></li> </ul>	Comparison with competitors				Interlaboratory proficiency test
5)Pérez-Ramírez E. <i>et al.</i> (2020). <b>External quality assessment of Rift Valley fever diagnosis in 17 veterinary laboratories of the Mediterranean and Black Sea regions.</b> Plos One, 15(9), e0239478.	<ul style="list-style-type: none"> <li>external quality assessment performed using a panel of 10 sheep sera (6 positive and 4 negative); sera were analyzed using the ID Screen RIFT VALLEY FEVER Competition Multi-species, another commercial kit and VNT.</li> <li><b>Results:</b> -16 out of 17 laboratories reported correct results for all the samples showing excellent reproducibility between laboratories (kappa value = 1) -only one laboratory obtained one incorrect result (one sample was reported as doubtful instead of positive), also reaching a high reproducibility (kappa value = 0.82) -a laboratory providing two different datasets produced 100% of correct results with the ID Screen RIFT VALLEY FEVER COMPETITION MULTI-SPECIES kit and four false negative results with an alternative commercial kit. <b>The ID Screen RIFT VALLEY FEVER COMPETITION MULTI-SPECIES showed excellent performance, better than the other commercial kit.</b></li> </ul>					Interlaboratory proficiency test

<p>6)de Bronsvort B. <i>et al.</i> (2019). <b>Comparison of Two Rift Valley Fever Serological Tests in Cameroonian Cattle Populations Using a Bayesian Latent Class Approach.</b> <i>Front. Vet. Sci.</i> 6:258.</p>	<ul style="list-style-type: none"> <li>comparison of performances of the ID Screen RIFT VALLEY FEVER Competition Multi-species with <b>PRNT</b> in a serological study among <b>cattle</b> in naturally infected populations (n=1473).</li> <li><b>Results: both the ID Screen RIFT VALLEY FEVER Competition Multi-species and PRNT have comparable performances:</b> the ID Screen RIFT VALLEY FEVER COMPETITION MULTI-SPECIES had an estimated diagnostic sensitivity (Se) of 0.854 and specificity (Sp) of 0.986 using all the data and splitting the population by geographical region compared to 0.844 and 0.981 for the PRNT80.</li> </ul> <p><b><i>“This study supports the use of the ID Screen RIFT VALLEY FEVER COMPETITION MULTI-SPECIES as a relatively low-cost, easy-to-use surveillance tool for the African context (sic)”.</i></b></p>	Correlation with other techniques		Epidemiological study	Performance evaluation
<p>7)Lindahl J.F. <i>et al.</i> (2019). <b>A multiplex fluorescence microsphere immunoassay for increased understanding of Rift Valley fever immune responses in ruminants in Kenya.</b> <i>Journal of virological methods</i>, 269, 70-76.</p>	<ul style="list-style-type: none"> <li>Evaluation of a multiplexing fluorescence microsphere immunoassay (<b>FMIA</b>) for the detection of IgG and IgM antibodies in ruminant sera against the RVFV nucleocapsid Np, glycoprotein Gn, and non-structural protein NSs. <b>Sheep</b> and <b>cattle</b> sera were tested by FMIA, the ID Screen RIFT VALLEY FEVER COMPETITION MULTI-SPECIES, and another commercially available competitive ELISA.</li> <li><b>Results:</b> -FMIA revealed strong detection of RVFV antibodies against the Np, Gn and NSs antigen targets. -<b>FMIA Np and Gn targets showed to correlate well with the ID Screen RIFT VALLEY FEVER Competition Multi-species results for IgG detection</b> -there is a poor correlation of the FMIA to another commercial kit. This was supported by the lower sensitivity and specificity of the FMIA when compared to the other commercial kit versus the ID Screen RIFT VALLEY FEVER Competition Multi-species.</li> </ul>	Correlation with other techniques			
<p>8)Monaco F. <i>et al.</i> (2015). <b>First External Quality Assessment of Molecular and Serological Detection of Rift Valley Fever in the Western Mediterranean Region.</b> <i>PLoS ONE</i> 10(11): e0142129.</p>	<ul style="list-style-type: none"> <li>assessment of the diagnostic capacities of ten laboratories involved in the RVF surveillance was performed. For the serological diagnosis of RVF, each participant received a panel of 15 <b>ruminant</b> sera composed of 5 negative and 10 positive samples (5 samples were from RVFV vaccinated sheep (n = 4) and goats (n = 1) seropositive for IgG and 5 samples from 5 naturally infected springboks RVF seropositive for both IgG and IgM. To detect the RVF IgG antibodies, all participating laboratories used the ID Screen RIFT VALLEY FEVER Competition Multi-species, and all laboratories used the ID Screen RIFT VALLEY FEVER IgM Capture for the serological detection of IgM.</li> </ul>				External Quality Assessment

	<ul style="list-style-type: none"> <li><i>Results:</i> 9 out of the 10 laboratories reported 100% of correct results, whilst one laboratory reported all correct results except one false-positive.</li> </ul>					
<p>9) Kortekaas J. <i>et al.</i> (2012). <b>European ring trial to evaluate ELISAs for the diagnosis of infection with Rift Valley fever virus.</b> <i>Journal of Virological Methods</i>, 187(1), 177-181.</p>	<ul style="list-style-type: none"> <li>ring trial organized to evaluate Rift Valley fever virus (RVFV) ELISAs by European laboratories: 5 ELISAs, including the ID Screen RIFT VALLEY FEVER Competition Multi-species, were evaluated by six participants. Sera were derived from cattle or sheep and originated from either a RVFV endemic area, a RVFV-free area or from experimental infection studies.</li> <li><i>Results:</i> Diagnostic sensitivity and specificity of the ID Screen® ELISA were found to be 98% and 100%, respectively, by most labs, which was higher than for the other IgG / competition ELISAs tested.</li> </ul>					Interlaboratory proficiency test

## CATTLE

<p>10) Fafetine J. <i>et al.</i> (2025). <b>Zoonotic arbovirus infections in cattle in Mozambique with special reference to Crimean-Congo hemorrhagic fever virus (CCHFV) and rift valley fever virus (RVFV).</b> <i>Virology Journal</i>, 22(1), 185.</p>	<ul style="list-style-type: none"> <li>460 cattle sera were tested using the ID Screen RIFT VALLEY FEVER Competition Multi-species.</li> <li><i>Results:</i> seroprevalence was 28%.</li> </ul>					
<p>11) Mlacha J.A. <i>et al.</i> (2025). <b>Risk factors, patterns and seropositivity of inter-epizootic Rift Valley fever virus of cattle in northern Tanzania.</b> <i>bioRxiv</i>, 2025-10.</p>	<ul style="list-style-type: none"> <li>1627 cattle sera were tested using the ID Screen RIFT VALLEY FEVER Competition Multi-species.</li> <li><i>Results:</i> seroprevalence was 15%.</li> </ul>					
<p>12) Madzingira O. <i>et al.</i> (2024). <b>Seroprevalence of <i>Brucella spp.</i> and Rift Valley fever virus infections in communal pastoral cattle at the wildlife-livestock interface, Zambezi region, Namibia.</b> <i>Front. Vet. Sci.</i> 11:1489815.</p>	<ul style="list-style-type: none"> <li>371 cattle sera were tested using the ID Screen RIFT VALLEY FEVER Competition Multi-species.</li> <li><i>Results:</i> seroprevalence was 41%.</li> </ul>					

13)Bronsvoort B.M. <i>et al.</i> (2022). <b>A Cross-Sectional, Population-Based, Seroepidemiological Study of Rift Valley Fever in Cameroonian Cattle Populations.</b> Front. Vet. Sci. 9:897481.	<ul style="list-style-type: none"> <li>1458 cattle sera were tested using the ID Screen RIFT VALLEY FEVER Competition Multi-species in 2 study locations in Cameroon.</li> <li><i>Results:</i> seroprevalence was <ul style="list-style-type: none"> <li>-in Northwest Region: 6.5%</li> <li>-in Vina division: 8.2%.</li> </ul> </li> </ul>					
14)Hassan-Kadle A. <i>et al.</i> (2021). <b>Rift Valley fever and <i>Brucella spp.</i> in ruminants, Somalia.</b> BMC Veterinary Research, 17(1), 1-6.	<ul style="list-style-type: none"> <li>sera from 609 ruminants (201 cattle, 203 goats, and 205 sheep), were serologically screened using the ID Screen RIFT VALLEY FEVER Competition Multi-species.</li> <li><i>Results:</i> 2 cattle sera tested positive (prevalence 1%).</li> </ul>					
15)Métras R. <i>et al.</i> (2020). <b>Estimation of Rift Valley fever virus spillover to humans during the Mayotte 2018-2019 epidemic.</b> Proceedings of the National Academy of Sciences, 117(39), 24567-24574.	<ul style="list-style-type: none"> <li>1169 cattle sera were tested using the ID Screen RIFT VALLEY FEVER Competition Multi-species, results were related to human cases (RITFV-positive by RT-PCR).</li> <li><i>Results:</i> Both livestock and human surveillance data were used to parameterize a mathematical model.</li> </ul>					
16)Halawi A.A.D. <i>et al.</i> (2019). <b>Seroprevalence of Rift Valley fever in cattle of smallholder farmers in Kwilu Province in the Democratic Republic of Congo.</b> Tropical animal health and production, 51, 2619-2627.	<ul style="list-style-type: none"> <li>677 cattle sera were tested for anti-RVSV antibodies using immunofluorescent assay (IFA). Therefore, all IFA positive and intermediate and some negative sera were tested using the ID Screen RIFT VALLEY FEVER Competition Multi-species.</li> <li><i>Results:</i> -seroprevalence was 6.5% (44/677) -none of the IFA-negative samples tested positive on ELISA -38 animals (5.6%) were found positive for both IFA and ELISA -of the 7 samples with intermediate IFA results, 6 were confirmed positive on ELISA while 5 IFA-positive samples were ruled out negative on ELISA.</li> </ul> <p><b><i>It was interesting to note that most intermediate IFA results were confirmed by ELISA although a few IFA-positive samples were ruled out by ELISA. (sic)</i></b></p>	Correlation with other techniques				
17)Tshilenge G. <i>et al.</i> (2019). <b>Seroprevalence of Rift Valley fever virus in cattle in the Democratic Republic of the Congo.</b> Tropical animal health and production, 51, 537-543.	<ul style="list-style-type: none"> <li>1675 cattle sera were tested using the ID Screen RIFT VALLEY FEVER Competition Multi-species and the ID Screen RIFT VALLEY FEVER IgM Capture to detect both IgG and IgM. RT-PCR was used for the detection of nucleic acid of RVSV.</li> <li><i>Results:</i> -203 sera were IgG-positive (seroprevalence 12.37%) -among the IgG-positive samples screened for anti-RVSV IgM, only 1.47% (3/203) were IgM-positive -1 of the IgM-positive samples was positive by RT-PCR.</li> </ul>	Correlation with other techniques				



18)Alhaji N.B. <i>et al.</i> (2018). <b>Participatory survey of Rift Valley fever in nomadic pastoral communities of North-central Nigeria: The associated risk pathways and factors.</b> PLoS Negl Trop Dis 12(10): e0006858.	<ul style="list-style-type: none"> <li>97 cattle sera were tested using the ID Screen RIFT VALLEY FEVER Competition Multi-species</li> <li><i>Results:</i> seroprevalence was 11.3%.</li> </ul>					
19)Bazanow B.A. <i>et al.</i> (2018). <b>Preliminary serological investigation of Rift Valley fever in Poland.</b> Journal of Vector Borne Diseases, 55(4), 324-326.	<ul style="list-style-type: none"> <li>973 banked bovine sera were screened using the ID Screen RIFT VALLEY FEVER Competition Multi-species in an RVFV-free area.</li> <li><i>Results:</i> all sera tested negative.</li> </ul> <p><b>The ID Screen RIFT VALLEY FEVER Competition Multi-species shows 100% specificity in a RIFT-free area.</b></p>					Specificity data
20)Matiko M.K. <i>et al.</i> (2018). <b>Serological evidence of inter-epizootic/ interepidemic circulation of Rift Valley fever virus in domestic cattle in Kyela and Morogoro, Tanzania.</b> PLoS Negl Trop Dis 12(11): e0006931.	<ul style="list-style-type: none"> <li>356 sera s from the local breed of zebu cattle (<i>Bos indicus</i>) and <i>Bos indicus/Bos taurus</i> crossbreed were analyzed using the ID Screen RIFT VALLEY FEVER Competition Multi-species and the ID Screen RIFT VALLEY FEVER IgM Capture. A <b>PRNT</b> (Plaque Reduction Neutralizing Test) assay was performed in all competition ELISA-positive samples.</li> <li><i>Results:</i> -seroprevalence by the ID Screen RIFT VALLEY FEVER Competition Multi-species was 29.2% -8.4% of all cattle had RVFV IgM antibodies -when the 104 competition ELISA-positive samples were analyzed by PRNT, 89% (93/104) had RVFV-neutralizing antibodies.</li> </ul>	Correlation with other techniques				
21)Tshilenge G. <i>et al.</i> (2018). <b>Seroprevalence and virus activity of Rift Valley fever in cattle in eastern region of Democratic Republic of the Congo.</b> Journal of veterinary medicine, ID 4956378.	<ul style="list-style-type: none"> <li>450 cattle sera were analyzed using the ID Screen RIFT VALLEY FEVER Competition Multi-species and the ID Screen RIFT VALLEY FEVER IgM Capture. All the anti-RVF IgM samples were tested using RT-PCR.</li> <li><i>Results:</i> anti-RVF IgG prevalence was 6.22% -anti-RVF IgM prevalence was 1.8% -none of the positive anti-RVF IgM samples (n=8) was positive for viral RVFV RNA.</li> </ul>	Correlation with other techniques				
22)Fèvre M. <i>et al.</i> (2017). <b>An integrated study of human and animal infectious disease in the Lake Victoria crescent small-holder crop-livestock production system, Kenya.</b> BMC Infectious Diseases, Volume 17, Number 1, Page 1.	<ul style="list-style-type: none"> <li>983 cattle sera were tested using the ID Screen RIFT VALLEY FEVER Competition Multi-species.</li> <li><i>Results:</i> seroprevalence was 1,4%.</li> </ul>					

23)Umuhoza T. <i>et al.</i> (2017). <b>Seroprevalence of Rift Valley fever in cattle along the Akagera–Nyabarongo rivers, Rwanda.</b> Journal of the South African Veterinary Association 88, a1379.	<ul style="list-style-type: none"> <li>595 cattle sera were tested using the ID Screen RIFT VALLEY FEVER Competition Multi-species.</li> <li><i>Results:</i> seroprevalence was 16.8%.</li> </ul>					
24)Bagninbom J.M. (2016). <b>Séroprévalence et facteurs de risque de la fièvre de la vallée du rift chez les bovins dans les hautes terres du Cameroun.</b> caphavet.com/index.php/projets-2/memoires-et-theses/item/77-these-Dr-Bagninbom-Jean-Marc-Esmv-Ngaoundere-Cameroun.	<ul style="list-style-type: none"> <li>1498 bovine sera were tested using the ID Screen RIFT VALLEY FEVER Competition Multi-species.</li> <li><i>Results:</i> seroprevalence was 7.8%.</li> </ul>					

## SHEEP AND GOATS

25)Gahn M.C.B. <i>et al.</i> (2024). <b>Large-Scale Serological Survey of Crimean-Congo Hemorrhagic Fever Virus and Rift Valley Fever Virus in Small Ruminants in Senegal.</b> Pathogens 2024, 13, 689.	<ul style="list-style-type: none"> <li>sera from 1130 sheep and 997 goats were tested using the ID Screen RIFT VALLEY FEVER Competition Multi-species.</li> <li><i>Results:</i> seroprevalence was -in goats: 5.4% -in sheep: 3.5%.</li> </ul>					
26)Lysholm S. <i>et al.</i> (2022). <b>Crossing the Line: Seroprevalence and Risk Factors for Transboundary Animal Diseases Along the Tanzania-Zambia Border.</b> Front. Vet. Sci. 9:809128.	<ul style="list-style-type: none"> <li>977 sera from small ruminants were tested using the ID Screen RIFT VALLEY FEVER Competition Multi-species.</li> <li><i>Results:</i> seroprevalence was 2.76%.</li> </ul>					
27)Ebogo-Belobo J.T. <i>et al.</i> (2022). <b>Serological evidence of the circulation of the Rift Valley fever virus in sheep and goats slaughtered in Yaoundé, Cameroon.</b> Veterinary Medicine and Science, 8(5), 2114-2118.	<ul style="list-style-type: none"> <li>sera from 47 sheep and 144 goats were tested using the ID Screen RIFT VALLEY FEVER Competition Multi-species and the ID Screen RIFT VALLEY FEVER IgM Capture.</li> <li><i>Results:</i> -seroprevalence for IgG antibodies was in sheep: 6.4% in goats: 4.9% -no IgM detected in sheep and goats.</li> </ul>					

<p>28)Cichon N. <i>et al.</i> (2021). <b>Co-circulation of Orthobunyaviruses and Rift Valley Fever Virus in Mauritania, 2015.</b> Front. Microbiol. 12:766977.</p>	<ul style="list-style-type: none"> <li>458 sera from sheep and goats were analyzed using the ID Screen RIFT VALLEY FEVER Competition Multi-species, followed by <b>SNT</b> (seroneutralisation). Samples with positive and inconclusive results in ELISA were further tested with the ID Screen RIFT VALLEY FEVER IgM Capture. In case of divergent results in ELISA and SNT, a final assessment was performed with IFA. A parallel analysis was performed on 3 other orthobunyaviruses: Ngari virus (NRIV) Bunyamwera virus (BUNV) and Batai virus (BATV).</li> <li><b>Results:</b> the ID Screen RIFT VALLEY FEVER Competition Multi-species revealed 84 antibody-positive samples. 81/84 sera were confirmed seropositive using SNT, indicating a prevalence of 17.69%, and 22 samples revealed RVFV IgM antibodies (prevalence of 4.80%). -of the 81 specimens, 61 samples revealed antibodies against RVFV and at least against one of the three orthobunyaviruses.</li> </ul>	Correlation with other techniques				
<p>29)Fakour S. <i>et al.</i> (2021). <b>A serological and hematological study on Rift Valley fever and associated risk factors in aborted sheep at Kurdistan province in west of Iran.</b> Comparative Immunology, Microbiology and Infectious Diseases, 75, 101620.</p>	<ul style="list-style-type: none"> <li>182 sera from aborted sheep were analyzed using the ID Screen RIFT VALLEY FEVER Competition Multi-species. <b>IFA</b> was used to confirm positive samples.</li> <li><b>Results:</b> 3/182 sera were positive in both tests (prevalence 1.65%).</li> </ul> <p><b>The results of IFA were correlated with ID Screen RIFT VALLEY FEVER COMPETITION MULTI-SPECIES results.</b></p>	Correlation with other techniques				
<p>30)Alhaj M.S. <i>et al.</i> (2019). <b>The circulation of Rift Valley fever virus in sentinel animals in Saudi Arabia: a reterospective cohort study.</b> Basrah Journal of Veterinary Research, 18(2), 80-92.</p>	<ul style="list-style-type: none"> <li>sera from 330 sheep and goats were analyzed using the ID Screen RIFT VALLEY FEVER Competition Multi-species.</li> <li><b>Results:</b> seroprevalence was 10.09%.</li> </ul>					
<p>31)Poueme R. <i>et al.</i> (2019). <b>Seroprevalence and associated risk factors of Rift Valley fever in domestic small ruminants in the north region of Cameroon.</b> Veterinary medicine international.</p>	<ul style="list-style-type: none"> <li>sera from 355 goats and 325 sheep were analyzed using the ID Screen RIFT VALLEY FEVER Competition Multi-species; positive samples were tested using the ID Screen RIFT VALLEY FEVER IgM Capture.</li> <li><b>Results:</b> seroprevalence was -at animal level: 3.4%) -at herd-level: (24.6%) -no IgM detected.</li> </ul>					



32)Tshilenge G.M. <i>et al.</i> (2019). <b>Rift Valley fever virus in small ruminants in the Democratic Republic of the Congo.</b> Onderstepoort Journal of Veterinary Research 86(1), a1737.	<ul style="list-style-type: none"> <li>893 sera from sheep and goats from 7 provinces were analyzed using the ID Screen RIFT VALLEY FEVER Competition Multi-species and the ID Screen RIFT VALLEY FEVER IgM Capture.</li> <li><b>Results:</b> -seroprevalence values ranged: from 0.0% to 23.81% for goats from 0.0% to 37.11% for sheep -1 serum (1.85%) out of 54 that tested positive for IgG was found to be IgM-positive.</li> </ul>						
33)Mbotha D. <i>et al.</i> (2018). <b>Inter-epidemic Rift Valley fever virus seroconversions in an irrigation scheme in Bura, south-east Kenya.</b> Transboundary and Emerging diseases, 65(1), e55-e62.	<ul style="list-style-type: none"> <li>sera from 228 goats and 88 sheep were sampled during 6 visits and analyzed using the ID Screen RIFT VALLEY FEVER Competition Multi-species.</li> <li><b>Results:</b> at the beginning of the study, 229 animals of 247 were seronegative and an additional 65 seronegative animals of 69 were recruited during the study.</li> </ul>						
34)Blomström A-L. <i>et al.</i> (2016). <b>Seroprevalence of Rift Valley fever virus in sheep and goats in Zambezia, Mozambique.</b> Infection Ecology & Epidemiology.;6:10.3402/iee.v6.31343.	<ul style="list-style-type: none"> <li>368 sera from small ruminants were tested using the ID Screen RIFT VALLEY FEVER Competition Multi-species.</li> <li><b>Results:</b> seroprevalence was -in sheep: 44,2% -in goats: 25%.</li> </ul>						

## LARGE AND SMALL RUMINANTS

35)Jarra E. <i>et al.</i> (2025). <b>Sero-epidemiology of Rift Valley fever virus in ruminant livestock in The Gambia.</b> bioRxiv, 03.	<ul style="list-style-type: none"> <li>sera from 1416 cattle, 1101 goats and 1085 sheep were tested using the ID Screen RIFT VALLEY FEVER Competition Multi-species.</li> <li><b>Results:</b> seroprevalence was: -in cattle: 36.8% -in goats: 2.1% -in sheep: 4.3%.</li> </ul>						
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36)Sumaye R.D. <i>et al.</i> (2025). <b>Rift Valley Fever Virus Transmission During an Unreported Outbreak Among People and Livestock in South-Central Tanzania.</b> <i>Viruses</i> 2025, 17, 1329.	<ul style="list-style-type: none"> <li>sera from 2300 cattle, 1193 goats, and 956 sheep were tested using ID Screen RIFT VALLEY FEVER Competition Multi-species and the ID Screen RIFT VALLEY FEVER IgM Capture.</li> <li><i>Results:</i> -overall seroprevalence (IgG and/or IgM) was: in cattle:15.26% in goats: 3.77% in sheep: 5.65% -IgM seroprevalence was: in cattle1.24% in goats: 3.39% in sheep: 1.28%.</li> </ul>					
37)Kainga H. <i>et al.</i> (2022). <b>Seroprevalence and Associated Risk Factors of Rift Valley Fever in Livestock from Three Ecological Zones of Malawi.</b> <i>Pathogens</i> , 11, 1349.	<ul style="list-style-type: none"> <li>1523 sera from cattle, sheep, and goats were tested using the ID Screen RIFT VALLEY FEVER Competition Multi-species and the ID Screen RIFT VALLEY FEVER IgM Capture.</li> <li><i>Results:</i> -seroprevalence (IgG + IgM) was 17.14% for individual livestock 33.24% for livestock at herd-level 25.68% in sheep 21.35% in cattle 7.72% in goats.</li> </ul>	Correlation with other techniques				
38)Sado F.Y. <i>et al.</i> (2022). <b>Seroprevalence of Rift Valley fever virus in domestic ruminants of various origins in two markets of Yaounde, Cameroon.</b> <i>PLoS Negl Trop Dis</i> 16(8): e0010683.	<ul style="list-style-type: none"> <li>sera from 441 cattle, 168 goats, and 147 sheep were analyzed using the ID Screen RIFT VALLEY FEVER Competition Multi-species and the ID Screen RIFT VALLEY FEVER IgM Capture.</li> <li><i>Results:</i> -using the ID Screen RIFT VALLEY FEVER Competition Multi-species, seroprevalence was: 42.2% in cattle, 2.7% in sheep, and 2.4% in goats -IgM seroprevalence was: 1.1% in cattle, 1.4% in sheep, and 0% in goats.</li> </ul>	Correlation with other techniques				
39)Troupin C. <i>et al.</i> (2022). <b>Seroprevalence of brucellosis, Q fever and Rift Valley fever in domestic ruminants in Guinea in 2017–2019.</b> <i>BMC Veterinary Research</i> , 18(1), 64.	<ul style="list-style-type: none"> <li>sera from 463 cattle, 408 goats, and 486 sheep were tested using the ID Screen RIFT VALLEY FEVER Competition Multi-species. To confirm that RVFV was circulating and not another phlebovirus, a <b>sero-neutralisation test</b> was performed on the serum of 5 ELISA-positive and 3 negative cattle.</li> <li><i>Results:</i> -seroprevalence was in cattle: 16.4% in goats: 1% in sheep: 1% -neutralizing antibodies were detected only in the 5 ELISA-positive cattle.</li> </ul>	Correlation with other techniques				

<p>40)Wanjama J. <i>et al.</i> (2022). <b>Sero-Epidemiological Survey of Rift Valley Fever Virus in Ruminants in Nyandarua County, Kenya.</b> East African Agricultural and Forestry Journal, 86(1-2), 11-11.</p>	<ul style="list-style-type: none"> <li>sera from 164 cattle, 118 sheep, and 19 goats) were tested using the ID Screen RIFT VALLEY FEVER Competition Multi-species and the ID Screen RIFT VALLEY FEVER IgM capture.</li> <li><b>Results:</b> -seroprevalence was in cattle: 49.39% in sheep: 9.32% in goats: 10.53%. -94 IgG-positive sera samples were screened for IgM, and 3 cattle were found seropositive (prevalence 3.19%).</li> </ul>	Correlation with other techniques				
<p>41)Hassan-Kadle A. <i>et al.</i> (2021). <b>Rift Valley fever and <i>Brucella spp.</i> in ruminants, Somalia.</b> BMC Veterinary Research, 17(1), 1-6.</p>	<ul style="list-style-type: none"> <li>serum samples from 609 ruminants (201 cattle, 203 goats, and 205 sheep), were serologically screened using the ID Screen RIFT VALLEY FEVER Competition Multi-species</li> <li><b>Results:</b> 2 cattle sera tested positive (prevalence 1%).</li> </ul>					
<p>42)Wekesa F.C. <i>et al.</i> (2021). <b>Serological evidence of inter-epidemic circulation of Rift Valley fever virus in livestock in Kenya.</b> East African Agricultural and Forestry Journal, 85(3 &amp; 4), 13-13.</p>	<ul style="list-style-type: none"> <li>615 sera collected from Ovine, Caprine, and Bovine were analyzed using the ID Screen RIFT VALLEY FEVER Competition Multi-species, the ID Screen RIFT VALLEY FEVER IgM Capture and Serum Virus Neutralization Test (SNT). Samples that were competition-ELISA positive and capture IgM-ELISA negative were presumed to be IgG positive. All serum samples that were competition-ELISA positive were subjected to SNT and only samples that were IgM positive were subjected to qRT-PCR. Samples were also classified as positive if the results from SNT and/or qRT-PCR and Competition-ELISA and/or RVFV IgM Antibody Capture ELISA were consistently positive.</li> <li><b>Results:</b> -overall seropositivity IgG antibodies: 14%. -ovine are more susceptible to RVFV with anti-RVFV IgG, and IgM seropositivity rates of 17% and 23% respectively.</li> </ul>	Correlation with other techniques				
<p>43)Zouaghi K. <i>et al.</i> (2021). <b>First Serological Evidence of Crimean-Congo Hemorrhagic Fever Virus and Rift Valley Fever Virus in Ruminants in Tunisia.</b> Pathogens 2021, 10, 769.</p>	<ul style="list-style-type: none"> <li>699 sera from cattle, sheep, and goats were tested using the ID Screen RIFT VALLEY FEVER Competition Multi-species.</li> <li><b>Results:</b> seroprevalence was -in cattle: 3.3 % -in sheep: 2.6 % -in goats: 0%.</li> </ul>					

44)Durand B. <i>et al.</i> (2020). <b>Rift Valley fever in northern Senegal: A modelling approach to analyse the processes underlying virus circulation recurrence.</b> Plos Neglected Tropical Diseases, 14(6), e0008009.	<ul style="list-style-type: none"> <li>sera from both resident (n=168 sheep and 54 goats) and nomadic domestic herds (n=590 small ruminants and 70 cattle) were tested using the ID Screen RIFT VALLEY FEVER Competition Multi-species.</li> <li><b>Results:</b> seroprevalence was <ul style="list-style-type: none"> <li>-in resident herds:15.3%</li> <li>-in nomadic herds:12.4%.</li> </ul> </li> </ul>					
45)Mahmoud A. S. <i>et al.</i> (2018). <b>Rift Valley fever virus: a serological survey in Libyan ruminants.</b> Open Veterinary Journal, 8(2), 204-207.	<ul style="list-style-type: none"> <li>171 cattle and 686 small ruminant sera were tested using the ID Screen RIFT VALLEY FEVER Competition Multi-species.</li> <li><b>Results:</b> Antibodies against RVFV were not detected in any of the 857 sera tested.</li> </ul> <p><b>The ID Screen RIFT VALLEY FEVER COMPETITION MULTI-SPECIES. shows a specificity of 100% in a RIFT-free area.</b></p>					Specificity data
46)Fakour S. <i>et al.</i> (2017). <b>The first positive serological study on Rift Valley fever in ruminants of Iran.</b> Journal of vector borne diseases, 54(4), 348-352.	<ul style="list-style-type: none"> <li>sera from 288 ruminants (118 cattle, 142 sheep, and 28 goats) were tested using the ID Screen RIFT VALLEY FEVER Competition Multi-species and indirect immunofluorescence assay (IFA).</li> <li><b>Results:</b> -overall seroprevalence was: 1.74% <ul style="list-style-type: none"> <li>-seroprevalence in cattle was 1.7%</li> <li>-seroprevalence in sheep was: 2.11%</li> <li>-no goats tested positive</li> </ul> </li> <li><b>-results of IFA were correlated with the ID Screen RIFT VALLEY FEVER Competition Multi-species results.</b></li> </ul>	Correlation with other techniques				
47)Moiane B. <i>et al.</i> (2017). <b>High seroprevalence of Rift Valley fever phlebovirus in domestic ruminants and African Buffaloes in Mozambique shows need for intensified surveillance.</b> Infection ecology & epidemiology, 7(1), 1416248.	<ul style="list-style-type: none"> <li>sera from 1581 cattle, 1117 goats, 85 sheep, and 69 African buffaloes were tested using the ID Screen RIFT VALLEY FEVER Competition Multi-species.</li> <li><b>Results:</b> seroprevalence was <ul style="list-style-type: none"> <li>-in cattle: 37.3%</li> <li>-in African buffaloes: 30.4%</li> <li>-in sheep: 18.8%</li> <li>-in goats: 9.4%.</li> </ul> </li> </ul>					
48)Rissmann M. <i>et al.</i> (2017). <b>Evidence for enzootic circulation of Rift Valley fever virus among livestock in Cameroon.</b> Acta Tropica, 172, 7-13.	<ul style="list-style-type: none"> <li>serum samples of small ruminants (n=921) and cattle (n=1032) were analyzed using the ID Screen RIFT VALLEY FEVER Competition Multi-species, samples were additionally tested using an indirect IgG /Gn ELISA. Positive results of ELISA were confirmed with the serum neutralization test (SNT). In the case of negative SNT, samples were tested in <b>indirect immunofluorescence</b>. In those cases, the indirect immunofluorescence was determining the final assessment of the sample. All sera that were positive or inconclusive in the ID Screen RIFT VALLEY FEVER Competition Multi-species were</li> </ul>	Correlation with other techniques				

	<p>additionally tested in the ID Screen RIFT VALLEY FEVER IgM Capture for specific presence of IgM.</p> <ul style="list-style-type: none"> <li><i>Results:</i> -seroprevalence was 3.4% for small ruminants 13.5% for cattle -1 small ruminant and 3 cattle were found IgM positive.</li> </ul>					
49)Métras R. <i>et al.</i> (2016). <b>The Epidemiology of Rift Valley Fever in Mayotte: Insights and Perspectives from 11 Years of Data.</b> PLoS Negl Trop Dis 10(6): e0004783.	<ul style="list-style-type: none"> <li>retrospective and prospective serological surveys over 11 years in cattle and small ruminants (n= 5720) with the ID Screen RIFT VALLEY FEVER Competition Multi-species and the ID Screen RIFT VALLEY FEVER IgM Capture.</li> <li><i>Results: "This study showed the value of repeated serological testing to explain RVF population dynamics in this island population "(sic).</i></li> </ul>					
50)Kim H., Park J. <i>et al.</i> (2015). <b>Serological surveillance studies confirm the Rift Valley fever virus free status in South Korea.</b> Trop Anim Health Prod 47:1427–1430.	<ul style="list-style-type: none"> <li>2382 sera from goats and cattle were tested with the ID Screen RIFT VALLEY FEVER Competition Multi-species.</li> <li><i>Results:</i> all the samples were found negative.</li> </ul> <p><b>The ID Screen RIFT VALLEY FEVER COMPETITION MULTI-SPECIES. shows a specificity of 100% in a RIFT-free area.</b></p>					Specificity data
51)Roger M. <i>et al.</i> (2014). <b>Evidence for Circulation of the Rift Valley Fever Virus among Livestock in the Union of Comoros.</b> PLoS Neglected Trop.Dis.; 8(7): e3045.	<ul style="list-style-type: none"> <li>sera from 103 goats and 88 cattle were tested using the ID Screen RIFT VALLEY FEVER Competition Multi-species; positive samples were confirmed by VNT.</li> <li><i>Results:</i> -overall prevalence was 17.54% -all the positive samples were also positive by VNT.</li> </ul>	Correlation with other techniques				

## CAMELIDS

52)Kadja M.C. <i>et al.</i> (2025). <b>Seroprevalence of Rift Valley Fever Viruses Antibodies in Domestic Livestock in the Tahoua Region of Niger.</b> Veterinaria Italiana, 61(3).	<ul style="list-style-type: none"> <li>sera from 615 domestic ruminants (<b>cattle, goats, sheep and camels</b>) were tested using the ID Screen RIFT VALLEY FEVER Competition Multi-species.</li> <li><i>Results:</i> seroprevalence was -in camels: 36.56% -in cattle: 17.69% -in goats: 3.55% -in sheep: 3.37%.</li> </ul>					
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53)Megenas J.A. <i>et al.</i> (2024). <b>Seroprevalence and Co-Circulation of Rift Valley Fever Virus and West Nile Fever Virus in Livestock Population of Afar Region, Northeast Ethiopia.</b> Veterinary Medicine International, 2024(1), 8249077.	<ul style="list-style-type: none"> <li>sera from 224 <b>cattle</b>, 155 <b>camels</b>, 121 <b>goats</b>, 144 <b>sheep</b>, and 92 <b>donkeys</b> were tested using the ID Screen RIFT VALLEY FEVER Competition Multi-species and the ID Screen WNV Competition Multi-species.</li> <li><i>Results:</i> seroprevalence of RVFV and WNV co-circulation was <ul style="list-style-type: none"> <li>-in camels: 20.7%</li> <li>-in cattle: 7.14%</li> <li>-in donkeys: 0%</li> <li>-in goats: 11.6%</li> <li>-in sheep: 3.5%.</li> </ul> </li> </ul>					
54)Eckstein S. <i>et al.</i> (2022). <b>Viral and Bacterial Zoonotic Agents in Dromedary Camels from Southern Tunisia: A Seroprevalence Study.</b> Microorganisms 2022, 10, 727.	<ul style="list-style-type: none"> <li>sera from 500 <b>dromedary camels</b> were tested using the ID Screen RIFT VALLEY FEVER Competition Multi-species.</li> <li><i>Results:</i> seroprevalence was 1%.</li> </ul>					
55)Adamu A.M. <i>et al.</i> (2021). <b>Risk factors for Rift Valley fever virus seropositivity in one-humped camels (<i>Camelus dromedarius</i>) and pastoralist knowledge and practices in Northern Nigeria.</b> One health, 13, 100340.	<ul style="list-style-type: none"> <li>720 sera from <b>camels</b> were tested using the ID Screen RIFT VALLEY FEVER Competition Multi-species.</li> <li><i>Results:</i> seroprevalence was 19.9%.</li> </ul>					
56)Cosseddu G.M. <i>et al.</i> (2021). <b>Sero-surveillance of emerging viral diseases in camels and cattle in Nouakchott, Mauritania: an abattoir study.</b> Tropical Animal Health and Production, 53, 1-6.	<ul style="list-style-type: none"> <li>159 sera from <b>camels</b> and 118 sera from <b>cattle</b> were analyzed using the ID Screen RIFT VALLEY FEVER Competition Multi-species.</li> <li><i>Results:</i> seroprevalence was <ul style="list-style-type: none"> <li>-in camels: 45%</li> <li>-in cattle: 16%.</li> </ul> </li> </ul>					
57)Kalthoum S. <i>et al.</i> (2021). <b>Risk based serological survey of Rift Valley fever in Tunisia (2017–2018).</b> Heliyon, 7(9).	<ul style="list-style-type: none"> <li>173 sera from <b>camels</b> were analyzed using the ID Screen RIFT VALLEY FEVER Competition Multi-species.</li> <li><i>Results:</i> no seropositive camels were detected.</li> </ul> <p><b>The ID Screen RIFT VALLEY FEVER COMPETITION MULTI-SPECIES. shows a specificity of 100% in a RIFT-free area.</b></p>					Specificity data
58)Musa H.I. <i>et al.</i> (2021). <b>Survey of antibodies to Rift Valley fever virus and associated risk factors in one-humped camels (<i>Camelus dromedarius</i>) slaughtered in Maiduguri abattoir, Borno State, Nigeria.</b> Tropical Animal Health and Production, 53, 1-8.	<ul style="list-style-type: none"> <li>92 sera from <b>camels</b> were analyzed using the ID Screen RIFT VALLEY FEVER Competition Multi-species.</li> <li><i>Results:</i> seroprevalence was 20.7%.</li> </ul>					



59)Selmi R. <i>et al.</i> (2020). <b>First serological evidence of the Rift Valley fever <i>Phlebovirus</i> in Tunisian camels.</b> Acta tropica, 207, 105462.	<ul style="list-style-type: none"> <li>47 sera from <b>camels</b> were analyzed using the ID Screen RIFT VALLEY FEVER Competition Multi-species.</li> <li><i>Results:</i> seroprevalence was 37%.</li> </ul>					
60)Gür S. <i>et al.</i> (2017). <b>The first serological evidence for Rift Valley fever infection in the camel, goitered gazelle and Anatolian water buffaloes in Turkey.</b> Tropical Animal Health and Production 49: 1531-1535.	<ul style="list-style-type: none"> <li>serological study in <b>camels, gazelles, and buffaloes</b>, using the ID Screen RIFT VALLEY FEVER Competition Multi-species.</li> <li><i>Results:</i> -all gazelles were negative -seroprevalence in camels was: 1.3% -seroprevalence in buffaloes was 8.5%.</li> </ul>					
61)Hassine T. <i>et al.</i> (2017). <b>Emerging vector-borne diseases in dromedaries in Tunisia: West Nile, Bluetongue, Epizootic Haemorrhagic Disease and Rift Valley fever.</b>	<ul style="list-style-type: none"> <li>118 sera from <b>dromedaries</b> were tested using the ID Screen RIFT VALLEY FEVER Competition Multi-species and <b>serum neutralization</b>.</li> <li><i>Results:</i> no evidence for circulation of RVF with both techniques.</li> </ul> <p><b>The ID Screen RIFT VALLEY FEVER COMPETITION MULTI-SPECIES showed excellent specificity in dromedaries in a RIFT-free area.</b></p>	Correlation with other techniques				Specificity data
62)Mroz C. <i>et al.</i> (2017). <b>Seroprevalence of Rift Valley fever virus in livestock during inter-epidemic period in Egypt, 2014/15.</b> BMC Vet Res;13(1):87.	<ul style="list-style-type: none"> <li>sera from 221 <b>camels</b>, 438 <b>sheep</b>, 26 <b>goats</b>, 188 <b>buffaloes</b>) were tested using the ID Screen RIFT VALLEY FEVER Competition Multi-species; additional confirmation of positive and inconclusive results in previous tests was performed with an in-house <b>immunofluorescence assay</b>; to detect recent infection, samples showing positive ELISA results (positive to IgG or IgM) in combination with negative IIFA results (detects only IgG) were further tested with the ID Screen RIFT VALLEY FEVER IgM Capture; investigation of neutralizing antibodies of the samples was carried out by <b>VNT</b>; sera were classified as positive when VNT alone or at least two assays produced consistent positive results.</li> <li><i>Results:</i> seroprevalence was -0% in goats -0,46% in sheep -3,17% in camels -5,85% in buffaloes.</li> </ul>	Correlation with other techniques				

63)Rissmann M. <i>et al.</i> (2017). <b>Serological and genomic evidence of Rift Valley fever virus during inter-epidemic periods in Mauritania.</b> <i>Epidemiology and Infection</i> , 145(5), 1058-1068.	<ul style="list-style-type: none"> <li>sera from <b>small ruminants, cattle, and camels</b> (1066 animals) were tested using the ID Screen RIFT VALLEY FEVER Competition Multi-species (and an-house IgG glycoprotein-based ELISA). Positive results were tested by <b>SNT</b> and <b>IFA</b>. Positive results for small ruminants and cattle tested using the ID Screen RIFT VALLEY FEVER IgM Capture, for camels using in-house indirect IgM Elisa for camelids.</li> <li><b>Results:</b> seroprevalence was <ul style="list-style-type: none"> <li>-in small ruminants: 3,8%</li> <li>-in cattle: 15,4% (1 bovine sample positive in IgM)</li> <li>-in camels: 32%.</li> </ul> </li> </ul>	Correlation with other techniques					
64)Abdallah M.M. <i>et al.</i> (2016). <b>A survey of Rift Valley fever and associated risk factors among the one-humped camel (<i>Camelus dromedaries</i>) in Sudan.</b> <i>Irish veterinary journal</i> , 69, 1-6.	<ul style="list-style-type: none"> <li>240 sera from <b>camels</b> were analyzed using the ID Screen RIFT VALLEY FEVER Competition Multi-species.</li> <li><b>Results:</b> seroprevalence was 9.6%.</li> </ul>						
65)Ould El Mamy A. <i>et al.</i> (2011). <b>Unexpected Rift Valley Fever Outbreak, Northern Mauritania.</b> <i>Emerging Infectious Diseases</i> . Vol. 17, No. 10.	<ul style="list-style-type: none"> <li>262 sera from <b>small ruminants</b> and 279 sera from <b>camels</b> were analyzed using the ID Screen RIFT VALLEY FEVER Competition Multi-species.</li> <li><b>Results:</b> seroprevalence was <ul style="list-style-type: none"> <li>-in camels: 33%</li> <li>-in small ruminants: 44%.</li> </ul> </li> </ul>						

## WILDLIFE AND OTHER SPECIES

66)Ramaroson H.S. <i>et al.</i> (2025). <b>Quantitative evaluation of dogs as sentinels for Rift Valley fever virus circulation in Madagascar.</b> <i>bioRxiv</i> , 2025-05.	<ul style="list-style-type: none"> <li>sera from 513 <b>dogs</b> and 135 <b>cattle</b> were tested using the ID Screen RIFT VALLEY FEVER Competition Multi-species.</li> <li><b>Results:</b> seroprevalence was <ul style="list-style-type: none"> <li>-in dogs: 4.5%</li> <li>-in cattle: 24.4%.</li> </ul> </li> </ul>						
67)Atuman Y.J. <i>et al.</i> (2022). <b>Serological evidence of antibodies to Rift Valley fever virus in wild and domestic animals in Bauchi State, Nigeria.</b> <i>Veterinary Medicine International</i> .	<ul style="list-style-type: none"> <li>sera samples from 106 wild animals (<b>waterbuck, wildebeest, eland, elephant, zebra, kudu, hartebeest</b>), 300 <b>cattle</b>, and 200 <b>horses</b> were analyzed using the ID Screen RIFT VALLEY FEVER Competition Multi-species.</li> <li><b>Results:</b> -overall seroprevalence in domestic and wild animals: 7.1% <ul style="list-style-type: none"> <li>-seroprevalence in cattle: 11.3%</li> <li>-seroprevalence in wildlife 8.5%</li> </ul> </li> </ul>						

	<ul style="list-style-type: none"> <li>-antibodies to RVFV were detected in <b>waterbuck</b> (10.4%) <b>wildebeest</b> (11.3%), <b>eland</b> (22.6%), and <b>elephant</b> (3.8%), and none of the sera from zebra, kudu, and hartebeest show detectable antibodies to RVFV</li> <li>-none of the sera from horses showed detectable antibodies to RVFV.</li> </ul>					
68)Chambaro H.M. <i>et al.</i> (2022). <b>An unusually long Rift Valley fever inter-epizootic period in Zambia: Evidence for enzootic virus circulation and risk for disease outbreak.</b> PLoS neglected tropical diseases, 16(6), e0010420.	<ul style="list-style-type: none"> <li>sera from 13 sheep, 259 goats, and 285 wild ungulates (including <b>buffaloes, impalas, warthogs, and hartebeests</b>) were tested using the ID Screen RIFT VALLEY FEVER Competition Multi-species.</li> <li><i>Results:</i> seroprevalence was <ul style="list-style-type: none"> <li>-in wild ungulates: 33.7%</li> <li>-in domestic ruminants: 5.6%.</li> </ul> </li> </ul>					
69)Gakuya F. <i>et al.</i> (2022). <b>Evidence of co-exposure with <i>Brucella spp</i>, <i>Coxiella burnetii</i>, and Rift Valley fever virus among various species of wildlife in Kenya.</b> PLOS Neglected Tropical Diseases, 16(8), e0010596.	<ul style="list-style-type: none"> <li>363 sera from 16 different wildlife species (199 <b>buffaloes</b>, 36 <b>giraffes</b>, 21 <b>zebras</b>, 7 <b>eland</b>s, 15 <b>oryxes</b>, 11 <b>waterbucks</b> 11 <b>gazelles</b>, 9 <b>impalas</b>, 8 <b>cheetahs</b>, 8 <b>elephants</b>, 8 <b>warthogs</b>, 7 <b>rhinos</b>, 5 <b>lions</b>, 4 <b>wildebeests</b>, 3 <b>hartebeest</b> and 1 <b>leopard</b>) were tested using the ID Screen RIFT VALLEY FEVER Competition Multi-species.</li> <li><i>Results:</i> -overall seroprevalence was 18.9% -antibodies were detected in <b>buffaloes, elands, wildebeest, impalas, oryxes, giraffes, elephants, rhinos, and cheetahs.</b></li> </ul>					
70)Ndengu M. <i>et al.</i> (2020). <b>Seroprevalence and associated risk factors of Rift Valley fever in cattle and selected wildlife species at the livestock/wildlife interface areas of Gonarezhou National Park, Zimbabwe.</b> Onderstepoort Journal of Veterinary Research, 87(1), 1-7.	<ul style="list-style-type: none"> <li>1011 sera from cattle and 161 sera from wild animals (111 <b>buffaloes</b>, 32 <b>impalas</b>, and 18 <b>kudus</b>) were tested using the ID Screen RIFT VALLEY FEVER Competition Multi-species.</li> <li><i>Results:</i> -seroprevalence was <ul style="list-style-type: none"> <li>in cattle: 46.7%</li> <li>in buffaloes: 11.7%.</li> <li>-all impala and kudu samples tested negative.</li> </ul> </li> </ul>					

<p>71)Rissmann M. <i>et al.</i> (2020). <b>Competency of amphibians and reptiles and their potential role as reservoir hosts for Rift Valley Fever Virus.</b> <i>Viruses</i>, 12(11), 1206.</p>	<ul style="list-style-type: none"> <li>30 <b>African common toads</b> and 32 <b>common agamas</b> were experimentally infected with 2 RVFV strains (MP12 and ZH501 strains) to test the competency of amphibians and reptiles for RVFV. Serology (on lymph and serum samples) was followed using the ID Screen RIFT VALLEY FEVER Competition Multi-species and VNT.</li> <li><i>Results:</i> no seroconversion was detected in toads; only a few RVFV MP-12-infected agamas (n = 2 out of 13) developed a faint neutralizing antibody response (at 16 and 21 dpi). 2 other agamas tested positive using the ID Screen RIFT VALLEY FEVER Competition Multi-species , seroconversion started at 16dpi.</li> </ul> <p><b>The study supposes that toads do not seem to play a role as a reservoir for RVFV at all; the role of agamas, albeit of minor importance, cannot be excluded.</b></p>				Experimental infection
<p>72)Métras R. <i>et al.</i> (2017). <b>Absence of Evidence of Rift Valley Fever Infection in <i>Eulemur fulvus</i> (Brown Lemur) in Mayotte During an Interepidemic Period.</b> <i>Vector Borne and Zoonotic Diseases</i>;17(5):358-360.</p>	<ul style="list-style-type: none"> <li>72 <b>brown lemurs</b> were tested using the ID Screen RIFT VALLEY FEVER Competition Multi-species.</li> <li><i>Results:</i> all sera were found negative using both tests.</li> </ul> <p><b>The ID Screen RIFT VALLEY FEVER COMPETITION MULTI-SPECIES showed excellent specificity in brown lemurs.</b></p>				Specificity data
<p>73)Wesula Lwande O. <i>et al.</i> (2015). <b>Spatio-temporal variation in prevalence of Rift Valley fever: a post-epidemic serum survey in cattle and wildlife in Kenya.</b> <i>Infection Ecology &amp; Epidemiology</i>, 5:1, 30106, iee.v5.30106.</p>	<ul style="list-style-type: none"> <li>177 <b>cattle</b> sera and 297 <b>wildlife</b> sera were tested using the ID Screen RIFT VALLEY FEVER Competition Multi-species.</li> <li><i>Results:</i> -all the cattle were negative -11,6% prevalence in wildlife species (<b>buffaloes, black rhinoceros, elephants, warthogs, and wildebeest tested positive</b>) -no detection in baboons, giraffes, vervet monkeys, and zebras.</li> </ul>				

## HUMANS

<p>74)Baragatti M. <i>et al.</i> (2025). <b>Seroepidemiological Reconstruction of Long-term Rift Valley Fever Virus Circulation in Nouakchott, Mauritania.</b> <i>The Journal of Infectious Diseases</i>, 231(5), e853-e861.</p>	<ul style="list-style-type: none"> <li>1319 <b>human</b> sera were tested using the ID Screen RIFT VALLEY FEVER Competition Multi-species; if a sample was positive, serum neutralization assay (<b>SNT</b>) was performed to verify the absence of cross-reactivity against Toscana virus (<b>TOSV</b>) and sandfly fever Sicilian virus (<b>SFSV</b>).</li> <li><i>Results:</i> -seroprevalence was 12% -all the positive samples were negative for TOSV and SFSV using SNT.</li> </ul>	Correlation with other techniques			EXclusivity data
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75)Marami D. <i>et al.</i> (2025). <b>Rift Valley fever virus and <i>Coxiella burnetii</i> infections among febrile patients, Eastern Ethiopia.</b> PLoS Negl Trop Dis 19(8): e0013375.	<ul style="list-style-type: none"> <li>sera from 415 randomly selected adult <b>febrile patients</b> were tested using the ID Screen RIFT VALLEY FEVER Competition Multi-species and the ID Screen RIFT VALLEY FEVER IgM Capture; PCR was used to detect RVF virus RNA.</li> <li><b>Results:</b> -seroprevalence was 5.2% -no IgM detected -no RIFT virus RNA detected.</li> </ul>					
76)Oakley R.B. <i>et al.</i> (2024). <b>Seroprevalence and risk factors for Q fever and Rift Valley fever in pastoralists and their livestock in Afar, Ethiopia: A One Health approach.</b> PLoS Negl Trop Dis 18(8): e0012392.	<ul style="list-style-type: none"> <li>sera from 232 <b>pastoralists</b> and 1377 <b>livestock</b> were tested using the ID Screen RIFT VALLEY FEVER Competition Multi-species.</li> <li><b>Results:</b> seroprevalence was -in pastoralists: 6.1% -in cattle: 8.3% -in goats: 2.7% -in sheep: 2.5% -in camels: 1.8%.</li> </ul>					
77)Situma S. <i>et al.</i> (2024). <b>Serological Evidence of Cryptic Rift Valley Fever Virus Transmission Among Humans and Livestock in Central Highlands of Kenya.</b> Viruses.2024, 16, 1927.	<ul style="list-style-type: none"> <li>sera from 1750 <b>humans</b> and 706 <b>livestock</b> (271 cattle, 89 sheep, 346 goats) were tested using the ID Screen RIFT VALLEY FEVER Competition Multi-species; a subset of 42 human samples consisting of 26 positive, 10 negative, and 6 indeterminate by ELISA testing were subjected to the RVF virus neutralization test (<b>VNT</b>).</li> <li><b>Results:</b> seroprevalence was -in humans: 2% (Cohen's kappa 0.64 and PABAK 0.66 demonstrated substantial <b>agreement between the ELISA and VNT assays</b>) -in cattle: 8.1% -in goats: 2.1% -in sheep: all animals were seronegative.</li> </ul>	Correlation with other techniques				
78)Johnson S.A.M. <i>et al.</i> (2023). <b>Evidence of Rift Valley Fever Virus Circulation in Livestock and Herders in Southern Ghana.</b> Viruses 2023, 15, 1346.	<ul style="list-style-type: none"> <li>sera from 253 <b>goats</b>, 246 <b>sheep</b>, 220 <b>cattle</b>, and 157 <b>herders</b>: 17.8% were tested using the ID Screen RIFT VALLEY FEVER Competition Multi-species. All animal samples that tested positive were then also tested using the ID Screen IgM Capture; all human samples were re-tested using an IgG sandwich ELISA to detect anti-RVFPV IgG and a IgM-capture ELISA to detect anti-RVFPV IgM.</li> <li><b>Results:</b> seroprevalence was: -in cattle: 24.1% -in sheep: 8.5% -in goats: 7.9% (with 0.6% seropositivity of IgM in livestock) -in herders: 17.8%, using both the ID Screen RIFT VALLEY FEVER Competition Multi-species and IgG sandwich ELISA (the 2 results agreed completely), and IgM seroprevalence of 8.3%.</li> </ul>	Correlation with other techniques				

<p>79)Muturi M. <i>et al.</i> (2023). <b>Ecological and subject-level drivers of interepidemic Rift Valley fever virus exposure in humans and livestock in Northern Kenya.</b> Scientific Reports, 13(1), 15342.</p>	<ul style="list-style-type: none"> <li>sera from 676 <b>humans</b> and 1864 <b>livestock</b> were tested using the ID Screen RIFT VALLEY FEVER Competition Multi-species; out of the 1864 livestock samples tested, a subset of 1103 samples were randomly selected for additional testing to detect the presence of IgM, using the ID Screen IgM Capture.</li> <li><b>Results:</b> -seroprevalence using the RIFT VALLEY FEVER Competition Multi-species was: in humans: 21.7% in livestock: 8.4% -IgM were detected in 0.4% of the livestock samples.</li> </ul>				
<p>80)Nyamota R. <i>et al.</i> (2023). <b>Seroprevalence of Brucella spp. and Rift Valley fever virus among slaughterhouse workers in Isiolo County, northern Kenya.</b> PLoS Negl Trop Dis 17(10): e0011677.</p>	<ul style="list-style-type: none"> <li>378 <b>slaughterhouse workers</b> were tested using the ID Screen RIFT VALLEY FEVER Competition Multi-species; seropositive samples were subjected to Virus Neutralization Test (<b>VNT</b>).</li> <li><b>Results:</b> -seroprevalence was 18.3% -out of the 69 samples that were positive for ELISA, 59 were confirmed to have anti-RVFP antibodies by VNT.</li> </ul>	Correlation with other techniques			
<p>81)Rwegoshola K. M. <i>et al.</i> (2023). <b>Seasonal Transmission Dynamics of Rift Valley Fever in Kilimanjaro Region, Tanzania.</b> East Africa Science, 5(1), 14-21.</p>	<ul style="list-style-type: none"> <li>sera from 446 <b>individuals</b> were tested using the ID Screen RIFT VALLEY FEVER Competition Multi-species.</li> <li><b>Results:</b> seroprevalence was 12.8%.</li> </ul>				
<p>82)de Glanville W.A. <i>et al.</i> (2022). <b>Inter-epidemic Rift Valley fever virus infection incidence and risks for zoonotic spillover in northern Tanzania.</b> PLoS Negl Trop Dis 16(10): e0010871.</p>	<ul style="list-style-type: none"> <li>sera from 558 <b>people</b> and 9476 <b>livestock</b> (3582 cattle, 3303 goats, and 2584 sheep) were screened using the ID Screen RIFT VALLEY FEVER Competition Multi-species.</li> <li><b>Results:</b> seroprevalence was -in livestock: 2.8% -in people: 8.2%.</li> </ul>				
<p>83)Oragwa A.O. <i>et al.</i> (2022). <b>Serologic evidence of silent Rift Valley fever virus infection among occupationally exposed persons in northern Nigeria.</b> The Journal of Infection in Developing Countries, 16(05), 881-887.</p>	<ul style="list-style-type: none"> <li>sera from 196 <b>individuals</b> comprising butchers (n = 121), abattoir/slaughterhouse workers (n = 55), and livestock keepers (n = 20) were screened using the ID Screen RIFT VALLEY FEVER Competition Multi-species.</li> <li><b>Results:</b> -overall seroprevalence was 19.9%.</li> </ul>				



84) Sindato C. <i>et al.</i> (2022). <b>Rift Valley fever seropositivity in humans and domestic ruminants and associated risk factors in Sengerema, Ilala, and Rufiji districts, Tanzania.</b> International Journal of Infectious Diseases, 122, 559-565.	<ul style="list-style-type: none"> <li>sera from 664 <b>humans</b>, 361 <b>cattle</b>, 394 <b>goats</b>, and 242 <b>sheep</b> were tested using the ID Screen RIFT VALLEY FEVER Competition Multi-species.</li> <li><b>Results:</b> seroprevalence was -in humans: 2.1% -in animals: 9.5%.</li> </ul>					
85) Ibrahim M. <i>et al.</i> (2021). <b>Sero-prevalence of Brucellosis, Q-fever and Rift Valley Fever in humans and livestock in Somali region, Ethiopia.</b> PLoS Neglected Tropical Diseases, 15(1), e0008100.	<ul style="list-style-type: none"> <li>sera from <b>humans</b> (n=190) and livestock (108 <b>cattle</b>, 141 <b>camels</b>, 252 <b>goats</b>, and 229 <b>sheep</b>) were tested using the ID Screen RIFT VALLEY FEVER Competition Multi-species.</li> <li><b>Results:</b> seroprevalence was -in humans: 13,2% -in cattle: 17.9% -in camels: 42.6% -in goats: 6.3% -in sheep: 7.4%.</li> </ul>					
86) Kumalija M.S. <i>et al.</i> (2021). <b>Detection of Rift Valley fever virus inter-epidemic activity in Kilimanjaro Region, North Eastern Tanzania.</b> Global health action, 14(1), 1957554.	<ul style="list-style-type: none"> <li>sera from 2986 <b>goats</b> and 266 <b>humans</b> were tested using the ID Screen RIFT VALLEY FEVER Competition Multi-species.</li> <li><b>Results:</b> seroprevalence was -in humans: 13.2% -in goats: 23.3%.</li> </ul>					
87) Budodo R. <i>et al.</i> (2020). <b>Serological evidence of exposure to Rift Valley, Dengue and Chikungunya Viruses among agropastoral communities in Manyara and Morogoro regions in Tanzania: A community Survey.</b> doi.org/10.1101/2020.01.16.908830.	<ul style="list-style-type: none"> <li>sera from 122 <b>humans</b> were tested using the ID Screen RIFT VALLEY FEVER Competition Multi-species.</li> <li><b>Results:</b> seroprevalence was 16.39%.</li> </ul>					
88) Opayele A. <i>et al.</i> (2018). <b>Rift Valley fever virus infection among livestock handlers in Ibadan, Nigeria.</b> Journal of Immunoassay and Immunochemistry, Vol. 39, No. 6, 609–621.	<ul style="list-style-type: none"> <li>sera from 265 <b>humans</b> were tested using the ID Screen RIFT VALLEY FEVER Competition Multi-species (in parallel with ID Screen® RIFT IgM ELISA)</li> <li><b>Results:</b> seroprevalence was 14,5%, with no IgM detected.</li> </ul>					

<p>89)Mohamed A.M. <i>et al.</i> (2014). <b>Seroepidemiological survey on Rift Valley fever among small ruminants and their close human contacts in Makkah, Saudi Arabia, in 2011.</b> Rev Sci Tech; 33(3): 903–915.</p>	<ul style="list-style-type: none"> <li>500 sera from <b>small ruminants</b> and 100 <b>human</b> sera were tested using the ID Screen RIFT VALLEY FEVER Competition Multi-species. All positive samples were retested using the ID Screen RIFT VALLEY FEVER IgM Capture.</li> <li><b>Results:</b> -seroprevalence was: 16,8% in animals 9% in humans -no sample was IgM positive.</li> </ul>					
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## EXPERIMENTAL /VACCINATION STUDIES

<p>90)Borrego B. <i>et al.</i> (2025). <b>The 40Fp8 vaccine strain is safe and protects pregnant ewes from a virulent RVFV challenge.</b> npj Vaccines, 10(1), 206.</p>	<ul style="list-style-type: none"> <li>evaluation of the attenuated <b>RVF-40Fp8 strain</b> in non-pregnant and pregnant <b>ewes</b>; challenge was performed using a virulent RVFV-56/74 strain; seroconversion was followed using the ID Screen RIFT VALLEY FEVER Competition Multi-species.</li> <li><b>Results:</b> immunized ewes seroconverted between 4- and 7-days post infection.</li> </ul>			Experimental study		
<p>91)Lubisi B.A. <i>et al.</i> (2023). <b>Experimental Infection of Domestic Pigs (<i>Sus scrofa</i>) with Rift Valley Fever Virus.</b> Viruses 2023, 15, 545.</p>	<ul style="list-style-type: none"> <li>experimental infection on pregnant <b>sows</b> using 2 RVF viruses (<b>M66/09</b> and <b>M21/10 variants</b>); seroconversion was followed using the ID Screen RIFT VALLEY FEVER Competition Multi-species.</li> <li><b>Results:</b> -M66/09 virus variant: all pregnant sows (n = 5) seroconverted from 14 days post infection (DPI) and remained positive until their humane euthanasia on different DPI; antibodies were detected in the offspring (n = 25; 75. 76%) of 3 of the sows which farrowed</li> <li>-M21/10 virus variant: half the sows (n = 2) and their offspring did not seroconvert, while the remaining sows (n = 2; 50%) and their piglets (n = 16; 43.2%) seroconverted from 4 DPI.</li> </ul>			Experimental study		
<p>92)Sindato C. <i>et al.</i> (2021). <b>Safety, Immunogenicity and Antibody Persistence of Rift Valley Fever Virus Clone 13 Vaccine in Sheep, Goats and Cattle in Tanzania.</b> Front. Vet. Sci. 8:779858.</p>	<ul style="list-style-type: none"> <li>a vaccine trial using <b>RVFV Clone 13</b> vaccine was conducted on 230 <b>sheep</b>, 230 <b>goats</b>, and 140 <b>cattle</b>. Animals were bled before vaccination and at days 15, 30, 60, 180, and 360 (+/- 10) post-vaccination to measure IgM and IgG antibody responses to RVFV using the ID Screen RIFT VALLEY FEVER Competition Multi-species and the ID Screen RIFT VALLEY FEVER IgM Capture.</li> <li><b>Results:</b> -by day 15 post-inoculation, the IgG seroconversion in vaccinated goats, cattle, and sheep was 27.0%, 20.0%, and 10.4%, respectively</li> </ul>			Vaccination monitoring		

	<p>-by day 30 post-inoculation, it was 75.0%, 74.1%, and 57.1% in vaccinated sheep, goats, and cattle, respectively</p> <p>-by day 60 post-inoculation, IgG seroconversion in sheep, goats, and cattle was 88.1%, 84.3%, and 64.60%, respectively</p> <p>-by day 180, the IgG seroconversion in sheep, goats, and cattle was 88.0%, 83.8%, and 66.1%, respectively</p> <p>-by day 360, the IgG seroconversion in sheep, goats, and cattle was 87.2%, 85.6%, and 66.1%, respectively</p> <p>-only five animals from the vaccinated group were RVFV IgM positive, which included four sheep and a goat.</p>					
93)Boumart Z. <i>et al.</i> (2019). <b>Safety and immunogenicity of a live attenuated Rift Valley Fever recombinant arMP-12ΔNSm21/384 vaccine candidate for sheep, goats and calves.</b> Vaccine, 37(12), 1642-1650.	<ul style="list-style-type: none"> <li>• evaluation of a <b>live attenuated recombinant RVFV vaccine</b> candidate in domestic ruminants. Immunogenicity among <b>sheep, goats, and calves</b> was followed using the ID Screen RIFT VALLEY FEVER Competition Multi-species and <b>VNT</b> (Virus Neutralization Test).</li> <li>• <i>Results:</i> detectable antibodies by day 7 post-vaccination among sheep, goats, and calves.</li> </ul>	Correlation with other techniques		Experimental vaccination		
94)Makoschey B. <i>et al.</i> (2016). <b>Rift Valley Fever Vaccine Virus Clone 13 Is Able to Cross the Ovine Placental Barrier Associated with Foetal Infections, Malformations, and Stillbirths.</b> PLoS Negl Trop Dis 10(3): e0004550.	<ul style="list-style-type: none"> <li>• <b>RVF vaccine virus clone 13</b> was tested in young <b>lambs</b> and pregnant <b>ewes</b>; the ID Screen RIFT VALLEY FEVER Competition Multi-species was used to follow antibody response.</li> <li>• <i>Results:</i> clone 13 virus induced RVFV antibody response in pregnant ewes 2 weeks post-inoculation and several pre-colostrum serum samples tested positive for RVFV-specific antibodies.</li> </ul>			Experimental vaccination		